# Activity of lactate dehydrogenase and superoxide dismutase in the circulation of patients with breast carcinoma

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#### **SUMMARY**

Background: A rapid increase in the turnover of malignant cells modulates the enzymes level in circulation and may be a Arch Oncol 2008;16(3-4):39-41. prognostic sign of disease progression. We evaluated the activity of serum lactate dehydrogenase and superoxide dismutase UDC: 618.19-006:616-089.168:577.11 in erythrocytes of breast cancer patients immediately after surgery and before adjuvant therapy.

Methods: Fourteen newly operated breast cancer patients were included in the study. Lactate dehydrogenase and superoxide dismutase activity were estimated using standard tests for evaluation of enzyme activity.

Results: Activity of lactate dehydrogenase was in normal range in all but one sample obtained from patient at clinical status Stanković", Belgrade, Serbia IIIB. Superoxide dismutase activity was elevated in 11 out of 14 patients, and was 2 to 8 fold higher compare to control values. Correspondence to: The highest activity of superoxide dismutase was found in samples of two patients at clinical status I.

Conclusion: Determination of lactate dehydrogenase and superoxide dismutase activity might be useful in clinical follow up of 21204 Sremska Kamenica, Serbia breast cancer patients.

Key words: Breast Neoplasms; Oxidative Stress; Lactate Dehydrogenases; Superoxide Dismutase; Postoperative Period

### INTRODUCTION

Breast cancer is the third most common cancer in women worldwide and accounts for the highest morbidity and mortality (1). Annually 910,000 new patients are diagnosed of breast cancer and 376,000 women die from the disease (2-4). According to the Registry for Malignant Neoplasms of Vojvodina, about 9000 women were diagnosed with breast cancer and 4800 died from the disease from 1995 to 2004 (5). Based on epidemiological studies, it has been established that risk factors for breast cancer include age, reproductive events (menarche, menopause, pregnancy), exogenous hormones (oral contraceptive and hormone replacement therapy), lifestyle risk factors (alcohol, diet, obesity) as well as genetic factors (high- and low-penetrance breast cancer susceptibility genes) among others (3,4). The increasing global incidence of breast cancer emphasizes the need to understand the various mechanisms involved in breast tumor genesis.

A rapid increase in the turnover of malignant cells modulates the enzymes level in the blood circulation (6). In addition, enzymatic changes reflect the overall changes in metabolism that occur in malignancy (7). Increased anaerobic glycolysis and activity of antioxidant enzymes were shown to be characteristics of cancer cell metabolism (6,8)

Lactate dehydrogenase (LDH) is released into the surrounding medium at an increased rate when cells replicate more rapidly (7,9). It was shown that LDH levels were higher in malignant than in normal tissue (10). LDH activity correlates with tumor mass or prognosis in human breast cancer. Metastatic breast cancer frequently exhibits elevated level of LDH as compared to normal breast tissue, but this observation is not consistent and still remains controversial (7,9)

Reactive oxygen species (ROS)-induced cell damage has been implicated in malignant transformation (11). The formation of free radicals is physiologically prevented or scavenged by host antioxidative defense mechanisms. Low levels of essential antioxidants in the circulation have been associated with an increased risk of cancer (12). Previously, impairment in antioxidant defense mechanisms has been demonstrated in a wide variety of malignancies including breast cancer (13-16). Basic and clinical studies showed that over-expression of manganese containing superoxide dismutase (MnSOD) may lead to changes of superoxide anion radical/ hydrogen peroxide  $(0_2)^{-1}$ H<sub>2</sub>O<sub>2</sub>) balance causing changes in the cell redox state. Changes in cell redox contains Mn, O<sub>2</sub> – Superoxide anion state affects signal transduction pathways that modulate cell proliferation (17-19). Over-expression of Cu,ZnSOD also significantly decreases growth and survival of breast cancer cells (14). Therefore, it is suggested that both MnSOD and Cu,ZnSOD can act as a tumor suppressors (14,20). On the other hand, some studies have reported correlation between high MnSOD level and invasiveness of breast cancer (20-23).

In the present study, we evaluated LDH and superoxide dismutase (SOD) Vojvodina, Sremska Kamenica activity in the circulation of patients with breast cancer. This is a preliminary report of results from the study on 14 newly diagnosed breast cancer patients obtained immediately after surgical intervention and before any adjuvant therapy.

### PATIENTS AND METHODS

Fourteen recently diagnosed women with breast cancer, median age 43 [30-49] from Institute of Oncology of Vojvodina, Serbia, were included in the study. They underwent surgical treatment and none of them was treated by hormonal, chemotherapy or radiotherapy. Patients' characteristics are shown in Table 1. Two out of 14 patients were at clinical stage I, five were classified as stage IIA, six as stage IIA, and one as stage IIIB. Informed consent was obtained from all the patients and local ethical board approved the study.

Blood samples were obtained from patients between the first and the second week after surgical treatment by venepuncture in tubes with EDTA. Plasma was separated by centrifugation at 3500 g for 10 min (Megafuge 1.0 R, Heraeus, Germany). After separation of plasma, the buffy coat was removed and the packed cells washed three times with physiologic saline, and centrifuged at 3500 g for 10 min. A known volume of erythrocytes was

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#### Abbreviations: Ca - Carcinoma

Cu. ZnSOD - Superoxide dismutase which contains Cu and Zn, EDTA -Ethylene-diammin-tetraascorbic acid. FAC - Therapeutic protocol for breast cancer patients, H<sub>2</sub>O<sub>2</sub> - Hydrogene peroxide, LDH - Lactate-dehydrogenase, MnSOD - Superoxide dismutase which radical, ROS - Reactive oxygene species, SOD - Superoxide dismutase

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Table 1. Characteristics of patients with breast carcinoma included in the study								
No	TNM class.	Clinical stage	Type of surgery	Hystological type				
1	$T_1N_{\scriptscriptstyle 0}M_{\scriptscriptstyle 0}$	I	Mastectomy	Lobular Carcinoma				
2	$T_1N_{\scriptscriptstyle 0}M_{\scriptscriptstyle 0}$	Ι	Breast conserving surgery	Lobular Carcinoma				
3	T,N1M₀	IIA	Breast conserving surgery	Lobular Carcinoma				
4	$T_{_2}N_{_0}M_{_0}$	IIA	Breast conserving surgery	Medullar Carcinoma				
5	$T_{\scriptscriptstyle 2}N_{\scriptscriptstyle 0}M_{\scriptscriptstyle 0}$	IIA	Mastectomy	Ductal Carcinoma				
6	$T_{z}N_{\scriptscriptstyle 0}M_{\scriptscriptstyle 0}$	IIA	Breast conserving surgery	Ductal Carcinoma				
7	$T_2N_0M_0$	IIA	Mastectomy	Ductal Carcinoma				
8	$T_2N_1M_0$	IIB	Mastectomy	Ductal Carcinoma				
9	$T_{\rm 2}N_{\rm 1}M_{\rm 0}$	IIB	Breast conserving surgery	Ductal Carcinoma				
10	$T_2N_1M_0$	IIB	Mastectomy	Ductal Carcinoma				
11	$T_{\rm z}N_{\rm 1}M_{\rm 0}$	IIB	Breast conserving surgery	Mucinous Carcinoma				
12	$T_{2}N_{1}M_{0}$	IIB	Mastectomy	Ductal Carcinoma				
13	$T_2N_1M_0$	IIB	Mastectomy	Ductal Carcinoma				

lysed with distillated water. Biochemical estimation for LDH was carried out immediately on ECTACHEM 250 apparatus according to original prescription by The Orthoclinical Jonhson & Jonhson (USA) producer.

Mastectomy

Ductal Carcinoma

Samples for SOD analysis were stored at -80°C until analysis (less than 1 month).

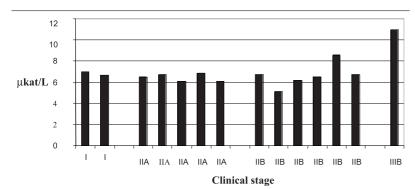


Figure 1. Activities of LDH in the serum of patients with breast cancer

14

T<sub>4</sub>N<sub>1</sub>M

IIIB

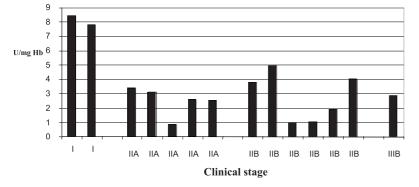


Figure 2. Activities of SOD in erythrocytes of patients with breast cancer

Erythrocyte SOD activity was assayed using the spectrophotometric indirect inhibition technique of Misra and Fridovich, based on the ability of SOD to inhibit auto-oxidation of adrenalin to adrenochrome at alkaline pH (24).

#### Statistical analysis

Data for biochemical analyses are expressed as individual and median values.

#### RESULTS

Individual values of LDH and SOD activity for all 14 breast cancer patients are shown in Table 2. Median values of LDH and SOD were 6.73 [5.12 - 11.00] ( $\mu$ kat/L) and 3.00 [0.87 - 8.45 ] (U/mg Hb) respectively.

LDH activity was in the normal range in all but one breast cancer patient. LDH activity in patient at clinical stage IIIB was approximately 2 fold higher compared to LDH values in lower stage samples (Table 2, Figure 1).

SOD was 2 to 8 fold increased in 11 patients in comparison to median physiological range for erythrocytes (27-29). Both patients at clinical stage I had the highest (~8 fold increased) level of SOD (Table 2, Figure 2). Significant individual differences in SOD activity were present in patients at IIA and IIB stage of disease.

#### Table 2. Activities of LDH and SOD in circulation of breast cancer patients

			LDH*	SOD*
No	Age	Clinical stage		
			(µkat/L)	(U/mg Hb)
1	30	I	6.98	8.45
2	47	I	6.70	7.81
3	39	IIA	6.50	3.38
4	43	IIA	6.76	3.12
5	43	IIA	6.12	0.87
6	35	IIA	6.89	2.59
7	45	IIA	6.07	2.54
8	43	IIB	6.76	3.76
9	32	IIB	5.12	4.96
10	43	IIB	6.23	0.98
11	49	IIB	6.49	1.02
12	44	IIB	8.54	1.88
13	39	IIB	6.75	4.00
14	35	IIIB	11.00	2.88

\*Ref. values: LDH: 5.22-10.30 µkat/L; SOD: 0.8-1 U/mg Hb (25-29).

## DISCUSSION

In this preliminary study, we investigated the activity of serum lactate dehydrogenase and superoxide dismutase in erythrocytes of patients with breast cancer. This investigation is a part of screening study dealing with effects of chemotherapy on antioxidative status and chromosomal damages in breast cancer patients. Results showed that only patient at clinical stage IIIB without distant metastases at the time of investigation had an increased LDH activity. Clinical follow-up of that patient showed relapse of primary disease with massive bone metastases and lethal outcome one year after ending of the chemotherapy. This result suggested that elevated level of LDH might be a prognostic sign of disease progression, which is in correlation with investigations of Seth et al. (7). Total LDH activity is elevated in the serum of about 65% cancer patients with metastases in the liver and in 20-60% cancer patients without hepatic metastases (25). LDH has also been found to correlate with tumor mass or prognosis in human breast cancer (7,30). Ryberg et al. reported that about 73% of breast cancer tissues had elevated serum LDH levels (30). According to Mehreen, LDH was not increased in patients without metastases (9), and we found normal LDH values in 13 samples in our screening investigation.

We found 2 to 8 fold increase of SOD in 11 patients in comparison to physiological level in erythrocytes (27-29). Both patients at clinical stage I had the highest (~8 fold increased) level of SOD.

The enzymatic antioxidant profile in the circulation of breast cancer patients shows an increase in SOD and GSH-Px activities compared with normal subjects, whereas the CAT activity showed a significant decrease compared with the respective controls (31). Decreased SOD in erythrocytes of patients with breast cancer has also been reported (27). Portakal et al. found that total SOD activity in breast tumor tissues was significantly higher than that in corresponding cancer-free tissues (32). Our findings are in accordance with that of Ray et al who reported that SOD activity was significantly raised in erythrocytes of breast cancer patients (16). Oxidative environment in newly established malignant phenotype may provoke oxidants to activate gene expression through the antioxidant responsive elements (33-35). It is supposed that over-expression of MnSOD, and probably, Cu,ZnSOD are important components of antioxidative defense through the process of malignant transformation (14). These facts might explain strongly enhanced SOD activity at lower stage of malignant disease that we found in two patients at clinical stage I.

In conclusion, lactate dehydrogenase and superoxide dismutase activity might be useful in clinical follow up of breast cancer patients.

#### Conflict of interest

We declare no conflicts of interest.

#### REFERENCES

- 1 Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics 2002. CA Cancer J Clin. 2005:55;74–108.
- 2 Kuzmiak M, Haberle S, Padungchaichote W, Zeng D, Cole E, Pisano EC. Insurance Status and the Severity of Breast Cancer at the Time of Diagnosis. *Acad Radiol.* 2008;15(10):1255-8.
- 3 Levil F, Te VC, Maspoli M, Randimbison L, Bulliard JL, Vecchia CL. Trends in breast cancer incidence among women under the age of forty. *BJC*. 2007;97:1013–4.
- 4 Stotter A, Bright N, Silcocks PB, Botha JL. Effect of improved data collection on breast cancer incidence and survival: reconciliation of a registry with a clinical database. *BMJ*. 2000;321:214.
- 5 Registry for Malign Neoplasms of Vojvodina, Oncology Institute of Vojvodina, 2004.
- 6 Yeh CC, Hou MF, Tsai SM, Lin SK and al. Superoxide anion radical, lipid peroxides and antioxidant status in the blood of patients with breast cancer. *Clin Chim Acta*. 2005;361:104-11.
- 7 Seth RK, Kharb S, Kharb DP. Serum biochemical markers in carcinoma breast. *Ind J Med Sci.* 2003;57;350-4.
- 8 Young CD, Anderson SM. Sugar and fat that's where it's at: metabolic changes in tumors. BCR. 2008;10(1):202-11.
- 9 Mehreen L, Khanam A. Evaluation of toxicities induced by chemotherapy in breast cancer patients. *Biomed Pharmacother*. 2005:59;524-7.
- 10 Kawamoto M. Breast cancer diagnosis by lactate dehydrogenase isoenzymes in nipple discharge. *Cancer Cytopathol.* 2008;73;1836–41.
- 11 Gonzalez RA. Free radicals, oxidative stress and DNA metabolism in human cancer. Cancer Invest. 1999:17;376–7.
- 12 Diplock AT. Antioxidant nutrients and disease prevention: an overview. Am J Clin Nutr. 1991:53;189–93.

- 13 Arivazhagan S, Kavitha K, Nagini S. Erythrocyte lipid peroxidation and antioxidants in gastric cancer. *Cell Biochem Funct*. 1997;15;15–8.
- 14 Nagini S, Saroja M. Circulating lipid peroxides and antioxidants as biomarkers of tumor burden in patients with oral squamous cell carcinoma. *J Biochem Mol Biol Biophys.* 2001:5;55–9.
- 15 Thangaraju M, Vijayalakshmi T, Sachdanandam P. Effect of tamoxifen on lipid peroxide and antioxidative system in postmenopausal women with breast cancer. *Cancer.* 1994;74:78–82.
- 16 Ray G, Batra S, Shukla NK, Deo S, Raina V, Ashok S, et al. Lipid peroxidation, free radical production and antioxidant status in breast cancer. *Breast Cancer Res Treat.* 2000:59;163–70.
- 17 Cullen JJ, Weydert C, Hinkhouse MM, Ritchie J, Domann FE, Spitz D, et al. The role of manganese superoxide dismutase in the growth of pancreatic adenocarcinoma. *Cancer Res.* 2003:63;1297-303.
- 18 Weydert CJ, Yhang Y, Sun W, Waugh TA, Teoh MLT, Andringa KK, et al. Increased oxidative stress created by adenoviral MnSOD or Cu,Zn SOD plus BCNU (1,3bis(chlorethyl) – 1- nitrosourea inhibits breast cancer growth. *Free Rad Biol Med.* 2008:44;856-67.
- 19 Weydert CJ, Waugh TA, Ritchie JM, Iyer KS, Smith JL, Li L, et al. Overexpression of manganese or copper-zinc superoxide dismutase inhibits breast cancer growth. *Free Rad Biol Med.* 2006:41;226-37.
- 20 Chuang TC, Liu JY, Lin CT, Tang YT, Ych MH, Chana SC, et al. Human manganese dismutase suppresses HER2/ncu-mediated breast cancer malignancy. *FEBS Lett.* 2007:581;4443-9.
- 21 Kattan Z, Minig V, Leroy P, Dauca M, Becuwe. P. Role of manganese superoxide dismutase on growth and invasive properties of human estrogen-independent breast cancer cells. *Breast Cancer Res Treat*. 2008;108;203-15.
- 22 Tsanou E, Ioachim E, Briasoulis E, Damala K, Charchanti A, Karavasilis V, et al. Immunohistochemical expression of superoxide dismutase (MnSOD) antioxidant enzyme in invasive breast carcinoma. *Hist Histopathol.* 2004:19;807-13.
- 23 Zhongkui L, Khaletziiy A, Wang J, Wong JWC, Oberley LW, Li JJ. Genes regulated in human breast cancer cells overexpressing manganese-containing superoxide dismutase. *Free Rad Biol Med.* 2001:30;260-67.
- 24 Misra HP, Fridovich I. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. J Biol Chem. 1972:247;3170–5.
- 25 Thangaraju M, Rameshbabu J, Vasavi H, Ilachezhian S, Vinitha R, Sachdanandam P. The salubrious effect of tamoxifen on serum marker enzymes, glycoproteines, and lysosomal enzymes level in breast cancer women. *Mol Cell Biochem.* 1998:185;85-94.
- 26 Ryberg M, Nielsen D, Osterlind K, Andersen PK, Skovsgaard T, Dombernovsky P. Increased pretreatment serum lactate dehydrogenase (LDH) is the most important determinant of central nervous system (CNS) metastases in patients with metastatic breast cancer. *EJC Supp.* 2003;1;133-4.
- 27 Abiaka C, Al-Avadi F, Al-Sayer H, Gulshan S, Behbehani A, Farghally M. Activities of erythrocyte antioxidant enzymes in cancer patients. J Clin Lab Anal. 2002;16;167-71.
- 28 Drabko K, Junak AB, Kowalczyk JR. Serum concentration of IL-2, IL-4, IL-10 and TNF-α in children with acute lymphoblastic leukemia – possible role of oxidative stress. *Centr Eur J Immunol.* 2008:33;146-9.
- 29 Djordjević N, Babić G, Marković SD, Ognjanović BI, Štajn AS, Žikić RV, et al. Oxidative stress and changes in antioxidative defense system in erythrocytes of preeclampsia in women. *Repr Tox.* 2008:25;213-8.
- 30 Ryberg M, Nielsen D, Osterlind K, Skovsgaard T, Dombernowsky P. Prognostic factors and long-term survival in 585 patients with metastatic breast cancer treated with epirubicin-based chemotherapy. *Ann Oncol.* 2001.12;81-7.
- 31 Gibananada R, Sanjay B, Nooton KS, Suryanarayan D, Vinod R, Seetharaman A, et al. Lipid peroxidation, free radical production and antioxidant status in breast cancer. *Breast Cancer Res Treat.* 2000: 59; 63–70.
- 32 Portakal O, Ozkaya O, Erden Inal M, Bozan B, Kosan M, Sayek J. Coenzyme Q10 concentrations and antioxidant status in tissues of breast cancer patients. *Clin Biochem.* 2000: 33;279-84.
- 33 Rushmore TH, Morton MR, Pickett CB. The antioxidant response element. J Biol Chem. 1991:266;11632-9.
- 34 Kinnula VL, Crapo JD. Superoxide dismutases in malignant cells and human tumors. *Free Rad Biol Med.* 2004;36;718-44.
- 35 Forman HJ. Use and abuse of exogenious  $H_2O_2$  in studies of signal transduction. *Free Rad Biol Med.* 2007:42;926-32.